

# Package ‘roots’

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**Title** Reconstructing Ordered Ontogenetic Trajectories

**Version** 1.0

**Description** A set of tools to reconstruct ordered ontogenetic trajectories from single cell RNAseq data.

**Depends** R (>= 3.0)

**Imports** animation (>= 2.4), rARPACK (>= 0.11-0), igraph (>= 1.0.0)

**License** GPL (>= 2)

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**VignetteBuilder** knitr

**NeedsCompilation** no

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<b>animPlot</b>	<i>Animation plot</i>
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## Description

Animation plot

## Usage

```
animPlot(data, ccm, delay = 0.1, darken = 1, lwd = 1, c.cex = 1,
         main = "", ...)
```

## Arguments

data	Dimensionality reduction plot
ccm	Dataframe of indices and momentums
delay	Delay between frames in seconds
darken	Passed to colGrad() function
lwd	Line width
c.cex	Size of points.
main	Plot title
...	Passed to plot() function

## Details

Animation plot Generates plot in base R that gradually updates giving the impression of an animation

## Value

Generates plot

## Author(s)

Wajid Jawaid

## Examples

```
## Not run:
xx <- animPlot(x, ccm)

## End(Not run)
```

---

animPlotGif                   *Generates a GIF animating*

---

## Description

Generates a GIF animation

## Usage

```
animPlotGif(data, ccm, delay = 0.1, darken = 1, lwd = 1, c.cex = 1,
  main = "", gif = "animation", img.name = "tempPlot", plot.par = NULL,
  point.col = "#333333", arrowLength = 0.1, ...)
```

## Arguments

data	Reduced dimensionality map to be used for visualisation
ccm	Dataframe of indices and momentums
delay	Delay between frames in seconds
darken	Passed to colGrad() function
lwd	Line width
c.cex	Size of points.
main	Title
gif	Name of movie
img.name	Name of temporary image files generated
plot.par	Passed to R base par() function
point.col	Colour of background points
arrowLength	Modify length of arrow
...	Passed to plot() function

## Details

Generates a GIF animation

## Value

Produces an animated GIF with given file name

## Author(s)

Wajid Jawaid

**Examples**

```
## Not run:
xx <- animPlotGif(x, ccm, gif = "animation")

## End(Not run)
```

**applyGaussianKernelwithVariableSigma**

*Apply Gaussian Kernel using Laleh Haghverdi's variable sigma*

**Description**

Apply Gaussian Kernel using Laleh Haghverdi's variable sigma

**Usage**

```
applyGaussianKernelwithVariableSigma(d2, rsigmas, csigmas = NULL)
```

**Arguments**

d2	Squared distance metric
rsigmas	Sigmas for cells in the rows
csigmas	Sigmas for cells in the columns

**Details**

Apply Gaussian Kernel using Laleh Haghverdi's variable sigma

**Value**

Returns matrix of same size as d2.

**Author(s)**

Wajid Jawaid

**Examples**

```
## Not run:
d <- applyGaussianKernelwithVariableSigma(dist, sigmas)

## End(Not run)
```

---

bgGeneNorm	<i>Normalise by background gene set</i>
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## Description

Normalise by background gene set

## Usage

```
bgGeneNorm(x, threshold = 0.05)
```

## Arguments

- |           |   |
|-----------|---|
| x         | Matrix to be normalised with cells in rows and genes in columns     |
| threshold | Default 0.05. The threshold below which a gene is deemed background |

## Details

Normalise by background gene set. Find background genes that are expressed at a lower percentage of the total library size per cell than 'threshold' parameter. These genes are used to calculate a normalisation factor.

## Value

Returns a normalised matrix of same dimensions as 'x'

## Author(s)

Wajid Jawaaid

## Examples

```
## Not run:  
normGenes <- bgGeneNorm(x)  
  
## End(Not run)
```

**calculateVariableSigmas**

*Calculates sigmas for a distance matrix*

---

**Description**

Calculates sigmas for a distance matrix

**Usage**

```
calculateVariableSigmas(d, knn)
```

**Arguments**

d	Square distance matrix with 0 diagonal
knn	Number of nearest neighbours to use for calculation

**Details**

Calculates sigmas for a distance matrix Using Laleh Haghverdi's method

**Value**

Returns a vector of sigmas

**Author(s)**

wj241

**Examples**

```
## Not run:  
sigmas <- calculateVariableSigmas(dist, 5)  
  
## End(Not run)
```

---

colGrad	<i>Generates a smooth colour gradient</i>
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---

**Description**

Generates a smooth colour gradient

**Usage**

```
colGrad(x, darken = 1)
```

**Arguments**

- |        |   |
|--------|---|
| x      | Number of colours required                                      |
| darken | Multiplication factor. Must be less than 1. Smaller the darker. |

**Details**

Generates a smooth colour gradient Goes from red to red/green to green to green/blue to blue to blu/red

**Value**

Returns vector of RGB colours

**Author(s)**

Wajid Jawaid

**Examples**

```
gradientColors <- colGrad(10)
```

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diffuseMat	<i>Generic diffusion function</i>
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**Description**

Generic diffusion function using automated individualised sigma calculation

**Usage**

```
diffuseMat(data, ndims = 20, nsig = 5, removeFirst = TRUE,  
useARPACK = TRUE, distfun = NULL, sigmas = NULL, sqdistmat = NULL)
```

## Arguments

data	Matrix of data with genes in rows and cells in columns.
ndims	Number of dimensions to return
nsig	For automatic sigma calculation
removeFirst	Default TRUE. Removes the first eigenvector
useARPACK	Default TRUE. Uses Arnoldi algorithm for eigenvector calculations
distfun	A different distance function that returns the <b>squared</b> distance
sigmas	Manually provide sigma
sqdistmat	<b>Squared</b> distance matrix. Give your own squared distance matrix.

## Details

Generic diffusion function using automated individualised sigma calculation.

A Gaussian kernel is applied to the chosen distance metric producing an  $n \times n$  square unnormalised symmetric transition matrix,  $A$ . Let  $D$  be an  $n \times n$  diagonal matrix with row(column) sums of  $A$  as entries. The density corrected transition matrix will now be:

$$D^{-1}AD^{-1}$$

and can be normalised:

$$B^{-1}D^{-1}AD^{-1}$$

where  $B$  is an  $n \times n$  diagonal matrix with row sums of the density corrected transition matrix as entries. The eigen decomposition of this matrix can be simplified by solving the symmetric system:

$$B^{-\frac{1}{2}}D^{-1}AD^{-1}B^{-\frac{1}{2}}R' = R'\lambda'$$

where  $R'$  is a matrix of the right eigenvectors that solve the system and  $\lambda'$  is the corresponding eigenvalue diagonal matrix. Now the solution of:

$$B^{-1}D^{-1}AD^{-1}R = R\lambda$$

in terms of  $R'$  and  $B^{-\frac{1}{2}}$  is:

$$B^{-1}D^{-1}AD^{-1}B^{-\frac{1}{2}}R' = B^{-\frac{1}{2}}R'\lambda'$$

and

$$R = B^{-\frac{1}{2}}R'$$

This  $R$  without the first eigen vector is returned as the diffusion map.

## Value

List output containing:

<i>values</i>	Eigenvalues, excluding the first eigenvalue, which should always be 1.
<i>vectors</i>	Matrix of eigen vectors in columns, first eigen vector removed.
<i>nconv</i>	Number of eigen vectors/values that converged.
<i>niter</i>	Iterations taken for Arnoldi algorithm to converge.
<i>nops</i>	Number of operations.
<i>val0</i>	1st eigen value - should be 1. If not be suspicious!
<i>vec0</i>	1st eigen vector - should be $n^{-\frac{1}{2}}$ , where n is the number of cells/samples.
<i>usedARPACK</i>	Predicates use of ARPACK for spectral decomposition.
<i>distfun</i>	Function used to calculate the squared distance.
<i>nn</i>	Number of nearest neighbours used for calculating sigmas.
<i>d2</i>	Matrix of squared distances, returned from distfun.
<i>sigmas</i>	Vector of sigmas. Same length as number of cells if individual sigmas were calculated, otherwise a scalar if was supplied.
<i>gaussian</i>	Unnormalised transition matrix after applying Gaussian.
<i>markov</i>	Normalised gaussian matrix.
<i>densityCorrected</i>	Matrix after applying density correction to markov.

### Author(s)

Wajid Jawaid

### References

- Haghverdi, L., Buettner, F., Theis, F.J., 2015. Diffusion maps for high-dimensional single-cell analysis of differentiation data. Bioinformatics 31, 2989–2998.
- Haghverdi, L., Büttner, M., Wolf, F.A., Buettner, F., Theis, F.J., 2016. Diffusion pseudotime robustly reconstructs lineage branching. Nat Meth 13, 845–848.
- Angerer, P., Haghverdi, L., Büttner, M., Theis, F.J., Marr, C., Buettner, F., 2016. destiny: diffusion maps for large-scale single-cell data in R. Bioinformatics 32, 1241–1243.

### Examples

```
## Not run:  
xx <- diffuseMat(x)  
  
## End(Not run)
```

diffuseProj

*Predicts diffusion map projection from new data points*

### Description

Predicts diffusion map projection from new data points

### Usage

```
diffuseProj(dm, x, data, distfun)
```

**Arguments**

<code>dm</code>	Output from <code>diffuseMat2</code> function
<code>x</code>	Matrix of new data points. Features in rows and cells in columns.
<code>data</code>	Original data used to generate diffusion map
<code>distfun</code>	A distance function that takes new data as first parameter and previous data as second variable returning a squared distance measure, with each sample in the rows and distance to previous data points in columns, e.g. <code>function(x, y) (1 - cor(x, y))^2</code> .

**Details**

Predicts diffusion map projection from new data points

**Value**

Returns a matrix with projected diffusion components.

**Author(s)**

Wajid Jawaid

**Examples**

```
## Not run:
y <- diffuseProj(xx, newData, oldData, function(z) (1-cor(z))^2)

## End(Not run)
```

**fastDist**

*Fast vectorised Euclidean distance calculator*

**Description**

Fast vectorised Euclidean distance calculator

**Usage**

```
fastDist(x, squared = FALSE)
```

**Arguments**

<code>x</code>	Matrix with vectors in columns.
<code>squared</code>	Will not perform the square root, i.e. will return the squared ‘L2-norm’.

**Details**

Calculates Euclidean distances between vectors arranged as columns in a matrix.

**Value**

Returns a matrix of pairwise distances

**Author(s)**

Wajid Jawaid

**Examples**

```
## Not run:  
dist <- fastDist(x)  
  
## End(Not run)
```

---

filterGenes

*Filter genes*

---

**Description**

Filter genes

**Usage**

```
filterGenes(x, mu = 0.01, cv = 2, fano = FALSE)
```

**Arguments**

x	Matrix to be normalised with cells in rows and genes in columns
mu	Mean threshold
cv	Coefficient of variation or Fano factor threshold.
fano	Default TRUE. Predicate treat CV as Fano factor or CV

**Details**

Filter genes Filter genes by mean and either coefficient of variation, cv or Fano factor.

**Value**

Returns a filtered matrix with same number of cells but fewer genes than 'x'

**Author(s)**

Wajid Jawaid

**Examples**

```
## Not run:
expressionGenesFiltered <- filterGenes(x)

## End(Not run)
```

**findLouvain***Louvain clustering on transition matrix***Description**

Louvain clustering on transition matrix

**Usage**

```
findLouvain(mkv)
```

**Arguments**

<code>mkv</code>	Transition matrix
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**Details**

Louvain clustering on transition matrix

**Value**

Returns a list with graph, dataframe and community object

**Author(s)**

Wajid Jawaid

**Examples**

```
## Not run:
xx <- findLouvain(mkv)
xx$cll

## End(Not run)
```

---

fnc	<i>Find next cell function</i>
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---

**Description**

Find next cell function

**Usage**

```
fnc(rdmap, tm, curInd, mom = NULL, momAdj = 0.5, w1 = exp(1), w2 = 1,
varEst = 10)
```

**Arguments**

rdmap	reduced dimensionality matrix with cells in rows and dims in columns
tm	Transition matrix
curInd	Current state on tm
mom	Current momentum vector
momAdj	Weighting to adjust momentum. From 0-1. Lower numbers make smaller adjustment to momentum vector.
w1	Parameter - Base used for modifying of tm probs.
w2	Parameter - Multiplifaction factor used for modifying tm probs.
varEst	Number of alternatives to sample for estimating variance.

**Details**

Find next cell function. Transition probabilities are modified by calculating the cosine of the angle between the current momentum vector and the vector on the rdmap required for each transition. The transisiton probability is adjusted by multiplying by  $w1^{w2 * (\cosine\_angle)}$  and then normalising.

**Value**

Returns index of new cell and new momentum vector

**Author(s)**

Wajid Jawaid

**Examples**

```
## Not run:
nextCell <- fnc(rdmap, tm, curInd)

## End(Not run)
```

<code>getTraj</code>	<i>Find a plausible developmental journey</i>
----------------------	---

### Description

Return a plausible developmental journey

### Usage

```
getTraj(rdmap, tm, sourceCellInds, terminalCellsInd = NULL, momAdj = 0.5,
w1 = exp(1), w2 = 1, simLen = 50, sim.seed = NULL, varEst = 10)
```

### Arguments

<code>rdmap</code>	reduced dimensionality matrix with cells in rows and dims in columns
<code>tm</code>	Transition matrix
<code>sourceCellInds</code>	Starting sell indices
<code>terminalCellsInd</code>	Terminal cell indices
<code>momAdj</code>	Weighting to adjust momentum. From 0-1. Lower numbers make smaller adjustment to momentum vector.
<code>w1</code>	Parameter - Base used for modifying of tm probs.
<code>w2</code>	Parameter - Multiplifaction factor used for modifying tm probs.
<code>simLen</code>	Maximum number of allowable transisitons
<code>sim.seed</code>	Random seed for reproducibility
<code>varEst</code>	Number of alternatives to sample for estimating variance.

### Details

Return a plausible developmental journey

### Value

Returns a data.frame of ordered indices and momentums

### Author(s)

Wajid Jawaid

### Examples

```
## Not run:
traj <- getTraj(rdmap, tm, startCells, terminalCells)

## End(Not run)
```

---

goggles	<i>View single cell dataset</i>
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---

## Description

View single cell dataset

## Usage

```
goggles(x, pcaDims = 90, nsig = 5, dmat = NULL, mkv = NULL,  
plotDims = 2, kernSq = 2, ...)
```

## Arguments

x	Matrix with cells in rows and gene in columns
pcaDims	Number of PCA dimensions to keep for distance measure
nsig	Number of significant neighbours to keep for Gaussian kernel
dmat	Optional. Give your own distance matrix
mkv	Optional. Give your own markov matrix.
plotDims	Default 2. Number of dimensions to plot
kernSq	Factor to tighten kernel - operates on sigmas.
...	Additonal parameters not currently in use

## Details

View single cell dataset

## Value

A list of 1, dimensionality reduced data.frame; clust, returned from louvainClust(); adj, Sparse, pruned adjacency matrix; dmat, distance matrix; pca, PCA reduced matrix. sparse, diagnostics on adj prior to applying sparseMarkov().

## Author(s)

Wajid Jawaid

## Examples

```
## Not run:  
xx <- goggles(x)  
plot(xx$1)  
  
## End(Not run)
```

---

**sparseMarkov**                  *Make markov matrix sparse*

---

### Description

Make markov matrix sparse

### Usage

`sparseMarkov(mkv, knn)`

### Arguments

<code>mkv</code>	Markov matric
<code>knn</code>	Number of nearest neighbours. See above.

### Details

Make markov matrix sparse Choose knn as the maximum number of similar cells are likely to exist in your dataset.

### Value

Markovian sparse matrix.

### Author(s)

Wajid Jawaid

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