

# Package ‘pGRN’

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**Type** Package

**Title** Single-Cell RNA Sequencing Pseudo-Time Based Gene Regulatory Network Inference

**Version** 0.3.5

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**Description** Inference and visualize gene regulatory network based on single-cell RNA sequencing pseudo-time information.

**License** MIT + file LICENSE

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.2.3

**VignetteBuilder** knitr, rmarkdown

**Depends** R (>= 2.10),

**Imports** bigmemory, doParallel, dtw, foreach, ggplot2, ggraph, igraph, lmtest, proxy, tidygraph, visNetwork, future

**Suggests** knitr, webshot, rmarkdown,

**NeedsCompilation** no

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## R topics documented:

data_transform . . . . .	2
get_dtw_dist_bidirectional . . . . .	3
get_dtw_dist_mat . . . . .	3
get_networks . . . . .	4
matrix2adj . . . . .	5
module_networks . . . . .	6
pGRN . . . . .	7

pGRNDB . . . . .	8
plot_network . . . . .	9
plot_network_i . . . . .	9
run_dtw . . . . .	10
run_granger_test . . . . .	11
slideWindows . . . . .	12
<b>Index</b>	<b>13</b>

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**data\_transform***Pseudotime based Expression Data Transformation***Description**

Based on single-cell pseudotime information, get the sliding window average expression, and then standard normalize the expression for each gene

**Usage**

```
data_transform(data, pseudotime, slide_window_size = 100, slide_step_size = 50)
```

**Arguments**

<b>data</b>	expression matrix data
<b>pseudotime</b>	list of pseudotime
<b>slide_window_size</b>	sliding window size
<b>slide_step_size</b>	sliding window step size

**Value**

Transformed new matrix

**Examples**

```
data <- matrix(1,100,1000)
ptime <- seq(1:1000)
data_transform(data,
              ptime,
              slide_window_size=100,
              slide_step_size=50)
```

---

```
get_dtw_dist_bidirectional  
    Bidirectional DTW Distance
```

---

## Description

Get bidirectional DTW distance.

## Usage

```
get_dtw_dist_bidirectional(x, y)
```

## Arguments

x	list of x input
y	list of y input

## Value

numeric

## Examples

```
get_dtw_dist_bidirectional(c(1:1000),c(1:1000))
```

---

```
get_dtw_dist_mat      DTW distance matrix for all genes
```

---

## Description

Get DTW distance matrix for all genes using pseudotime based sliding window transformation, parallel computing allowed.

## Usage

```
get_dtw_dist_mat(  
  data,  
  ptime,  
  slide_window_size = 50,  
  slide_step_size = 25,  
  cores = 2  
)
```

**Arguments**

data	gene expression matrix (Gene * Cells)
ptime	pseudotime matched with the column cells of the gene expression matrix
slide_window_size	sliding window size
slide_step_size	sliding window step size
cores	number of cores for parallel computing

**Value**

bidirectional DTW distance matrix

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
dtw_dist_matrix <- get_dtw_dist_mat(expression_matrix,
                                      pseudotime_list,
                                      cores=1)
```

get_networks	<i>Get the list of sub-networks</i>
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**Description**

Get sub-networks based on given adjacency data.frame input

**Usage**

```
get_networks(
  data,
  centrality_degree_mod = "out",
  components_mod = "weak",
  network_min_genes = 10
)
```

**Arguments**

data	adjacency data.frame
centrality_degree_mod	mode of centrality degree for popularity calculation
components_mod	mode of sub-network extraction methods
network_min_genes	minimal number of gene elements required for extracted sub-networks

**Value**

list of tabl\_graph objects

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
dtw_dist_matrix <- get_dtw_dist_mat(expression_matrix,
                                      pseudotime_list,
                                      cores=1)
adj_df <- matrix2adj(dtw_dist_matrix)
get_networks(adj_df, network_min_genes=5)
```

---

**matrix2adj**

*Convert distance matrix to adjacency dataframe*

---

**Description**

Convert distance matrix to adjacency dataframe for network construction.

**Usage**

```
matrix2adj(data, quantile_cutoff = 5)
```

**Arguments**

data	distance matrix
quantile_cutoff	an integer value (1-99) for quantile cutoff

**Value**

adjacency dataframe (with columns "from, to, distance,direction, similarity")

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
dtw_dist_matrix <- get_dtw_dist_mat(expression_matrix,
                                      pseudotime_list,
                                      cores=1)
adj_df <- matrix2adj(dtw_dist_matrix)
```

---

<b>module_networks</b>	<i>Get module level networks</i>
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### Description

Given a distance matrix, calculate gene modules based on hierarchical clustering method and then get module level networks

### Usage

```
module_networks(
  data,
  k = 10,
  quantile_cutoff = 10,
  centrality_degree_mod = "out",
  components_mod = "weak",
  network_min_genes = 10
)
```

### Arguments

data	distance matrix
k	number of gene clusters for module inference
quantile_cutoff	distance cutoff based on quantile(1-99) for edge identification
centrality_degree_mod	"in" or "out" for nodes popularity calculation
components_mod	"weak" or "strong" for sub-network components inference
network_min_genes	mininal number of genes required for a network

### Value

a list networks for each module

### Examples

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
dtw_dist_matrix <- get_dtw_dist_mat(expression_matrix,
                                      pseudotime_list,
                                      cores=1)
nets <- module_networks(dtw_dist_matrix,k=1,quantile_cutoff=50)
plot_network(nets[["module1"]])
```

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pGRN*pGRN: creates gene regulatory network based on single cell pseudo-time information*

---

## Description

Given single cell matrix and pseudotime, construct gene regulatory network (GRN)

## Usage

```
pGRN(
  expression_matrix,
  pseudotime_list,
  method = "DTW",
  slide_window_size = 20,
  slide_step_size = 10,
  centrality_degree_mod = "out",
  components_mod = "weak",
  network_min_genes = 10,
  quantile_cutoff = 5,
  order = 1,
  cores = 1
)
```

## Arguments

expression_matrix	expression matrix data
pseudotime_list	list of pseudotime
method	method for GRN construction: DTW, granger
slide_window_size	sliding window size
slide_step_size	sliding window step size
centrality_degree_mod	(for DTW method) mode of centrality degree for popularity calculation
components_mod	(for DTW method) mode of sub-network extraction methods (weak or strong)
network_min_genes	minimal number of gene elements required for extracted sub-networks
quantile_cutoff	an integer value (1-99) for quantile cutoff
order	(for granger method) integer specifying the order of lags to include in the auxiliary regression
cores	number of cores for parallel computing

**Value**

a list of `tbl_graph` objects

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime

# try DTW method
nets <- pGRN(expression_matrix,
               pseudotime_list,
               method= "DTW",
               quantile_cutoff=50,
               cores=1)
plot_network(nets[[1]])

# plot the network interactively
plot_network_i(nets[[1]])
```

`pGRNDB`

*pGRN example data*

**Description**

A list with expression dataframe and pseudotime dataframe

**Usage**

`pGRNDB`

**Format**

**pGRNDB:**

A list with items `expression` and `ptime`

**expression** data frame of single cell expression

**ptime** pseudotime of the single cells ...

**Source**

`pGRN`

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plot_network	<i>Plot stationary network</i>
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**Description**

Plot stationary network through ggraph

**Usage**

```
plot_network(graph, ...)
```

**Arguments**

graph	a <code>tbl_graph</code> object
...	other parameters for <code>ggraph</code>

**Value**

`ggraph`

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
dtw_dist_matrix <- get_dtw_dist_mat(expression_matrix,
                                      pseudotime_list,
                                      cores=1)
nets <- module_networks(dtw_dist_matrix,k=1,quantile_cutoff=50)
plot_network(nets[["module1"]])
```

---

---

plot_network_i	<i>Plot interactive network</i>
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---

**Description**

Plot interactive network based on igraph layout input

**Usage**

```
plot_network_i(graph, save_file = NULL)
```

**Arguments**

graph	igraph layout object
save_file	file name of the saved file, not save if NULL

**Value**

```
visNetwork htmlwidget
```

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
dtw_dist_matrix <- get_dtw_dist_mat(expression_matrix,
                                       pseudotime_list,
                                       cores=1)
nets <- module_networks(dtw_dist_matrix,k=1,quantile_cutoff=50)
plot_network_i(nets[["module1"]])
```

---

**run\_dtw**

*Get network adjacency dataframe based on DTW method*

---

**Description**

Use DTW to calcuate gene-gene distance based on their expression and pseudotime

**Usage**

```
run_dtw(
  expression_matrix,
  pseudotime_list,
  slide_window_size = 50,
  slide_step_size = 25,
  quantile_cutoff = 5,
  cores = 1
)
```

**Arguments**

expression_matrix	expression matrix data
pseudotime_list	list of pseudotime
slide_window_size	sliding window size
slide_step_size	sliding window step size
quantile_cutoff	an integer value (1-99) for quantile cutoff
cores	number of cores for parallel computing

**Value**

adjacency dataframe (with columns "from, to, distance,direction, similarity")

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
adj_df <- run_dtw(expression_matrix,
                    pseudotime_list,
                    quantile_cutoff=50,
                    cores=1)
```

`run_granger_test`

*Use Granger-causality Test to get gene-gene regulatory relationship*

**Description**

Based on single-cell gene expression matrix and pseudotime, calculate Granger-causality Test based gene-gene regulatory relationship

**Usage**

```
run_granger_test(
  data,
  ptime,
  slide_window_size = 20,
  slide_step_size = 10,
  pvalue_cutoff = 0.01,
  order = 1,
  ...
)
```

**Arguments**

<code>data</code>	gene expression matrix (Gene * Cells)
<code>ptime</code>	pseudotime matched with the column cells of the gene expression matrix
<code>slide_window_size</code>	sliding window size
<code>slide_step_size</code>	sliding window step size
<code>pvalue_cutoff</code>	cutoff for the pvalue from transfer entropy test
<code>order</code>	integer specifying the order of lags to include in the auxiliary regression
<code>...</code>	other parameters for grangertest function in lmtest

**Value**

adjacency data frame

**Examples**

```
example_data <- pGRNDB
expression_matrix <- example_data[["expression"]]
pseudotime_list <- example_data[["ptime"]]$PseudoTime
gt_adj_df <- run_granger_test(expression_matrix, pseudotime_list)
```

---

**slideWindows**

*Sliding Window Average*

---

**Description**

Get sliding windows average values for given vector/list

**Usage**

```
slideWindows(data, window = 2, step = 1)
```

**Arguments**

<b>data</b>	list of expression
<b>window</b>	sliding window size
<b>step</b>	sliding window step size

**Value**

list/vector of sliding windows with average expression value

**Examples**

```
slideWindows(c(1:1000),window=200,step=100)
slideWindows(c(1:1000),window=100,step=50)
```

# Index

- \* **datasets**
  - pGRNDB, [8](#)
- data\_transform, [2](#)
- get\_dtw\_dist\_bidirectional, [3](#)
- get\_dtw\_dist\_mat, [3](#)
- get\_networks, [4](#)
- matrix2adj, [5](#)
- module\_networks, [6](#)
- pGRN, [7](#)
- pGRNDB, [8](#)
- plot\_network, [9](#)
- plot\_network\_i, [9](#)
- run\_dtw, [10](#)
- run\_granger\_test, [11](#)
- slideWindows, [12](#)